

Introduction

- Plankton are a group of primary producers that form the basis of all aquatic food webs and provide invaluable ecosystem services¹
- Plankton respond quickly to environmental changes and can threaten environmental and public health in the form of harmful blooms²
- Monitoring Plankton community composition is essential tool for coastal management
- Historically long-term plankton monitoring has been made difficult by limitations of traditional microscopy
- In recent years new tools have been developed to lower the amount of time and expertise required to enumerate plankton

Flow-Imaging-Microscopy

Flow-imaging microscopy methods involve the imaging, identification, and analysis of subsvisible particles suspended in a liquid medium¹

Advantages

- Rapid analysis of sample composition
- Automated generation of individual particle properties
- Images are preserved for future analysis
- Semi-automated particle classification capabilities

Disadvantages

- Inability to speciate most cell types
- Swapping hardware is required for capturing whole size spectra
- Particulate may obscure target particles
- Lack of standardized methodologies

Anatomy of a FlowCam

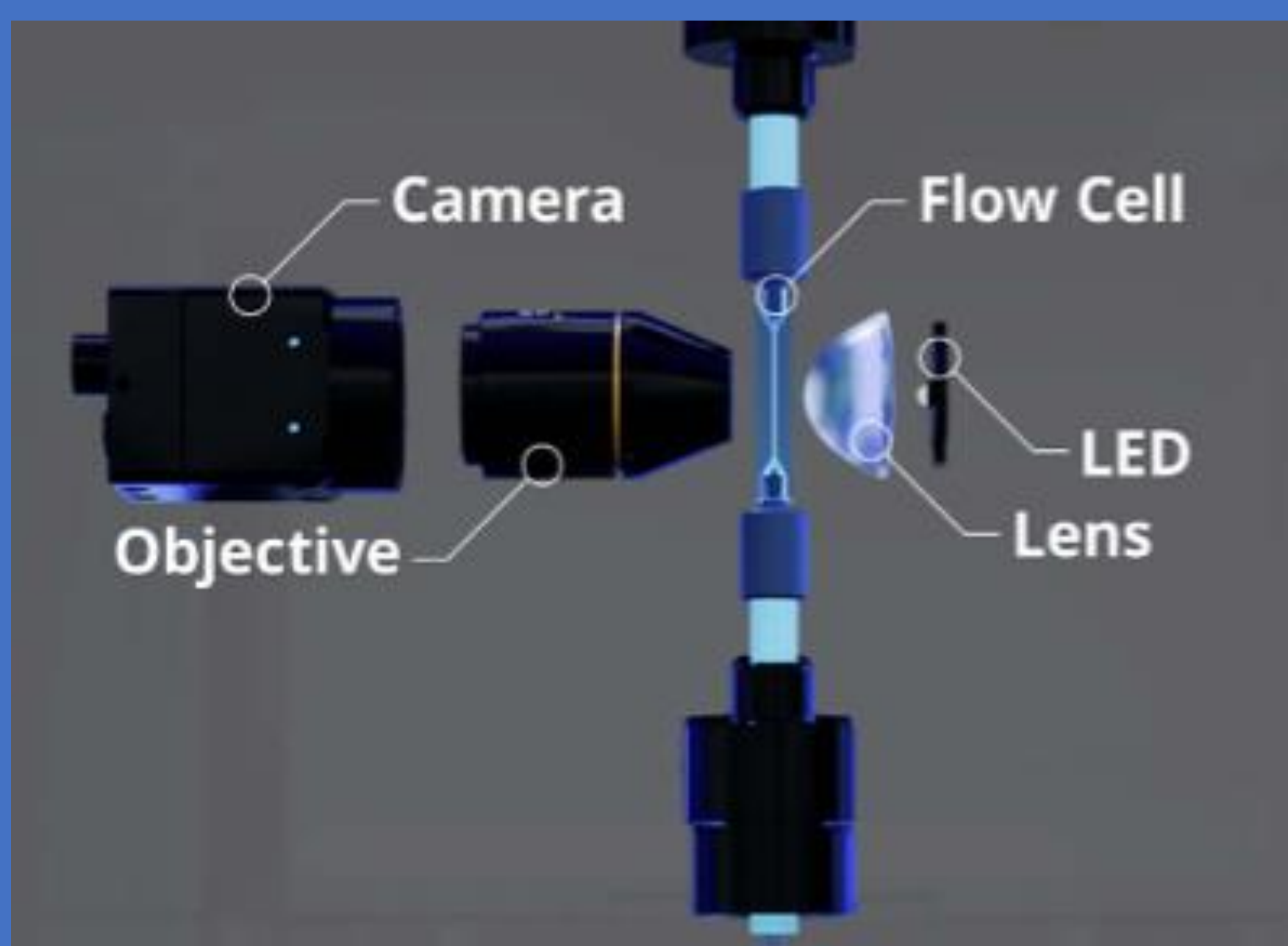
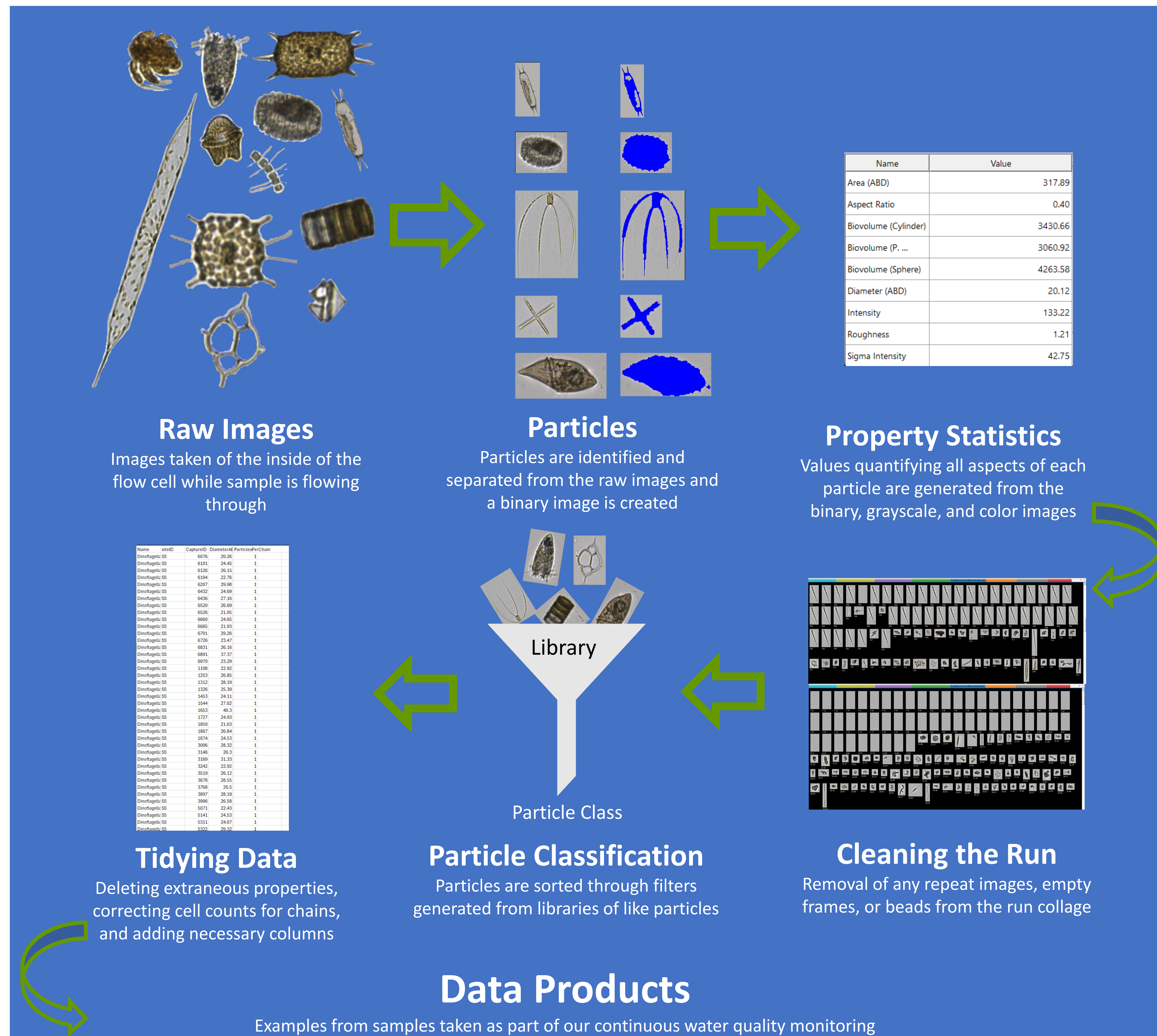


Fig. 1: Image illustrating the major components of a FlowCam flow-imaging microscope. (Credit: Yokogawa Fluid Imaging Technologies Inc.)³



| Name | Area | Aspect | Biovol | Diam | Inten | Rough | Sigma |
|------------|------|--------|--------|------|-------|-------|-------|
| FlowCam_01 | 6519 | 29.29 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_02 | 6351 | 24.45 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_03 | 6226 | 26.51 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_04 | 6284 | 22.76 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_05 | 6287 | 29.86 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_06 | 6452 | 24.69 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_07 | 6436 | 27.56 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_08 | 6529 | 26.69 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_09 | 6526 | 21.85 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_10 | 6660 | 24.65 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_11 | 6685 | 21.91 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_12 | 6751 | 29.26 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_13 | 6726 | 25.47 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_14 | 6821 | 26.31 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_15 | 6851 | 27.27 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_16 | 6970 | 23.29 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_17 | 1128 | 22.62 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_18 | 1273 | 26.85 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_19 | 1232 | 29.39 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_20 | 1326 | 25.39 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_21 | 1443 | 24.11 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_22 | 1544 | 27.02 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_23 | 1663 | 46.13 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_24 | 1727 | 24.63 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_25 | 1810 | 21.61 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_26 | 1867 | 26.44 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_27 | 1874 | 24.53 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_28 | 1906 | 26.32 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_29 | 1946 | 26.13 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_30 | 1939 | 32.33 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_31 | 1942 | 22.92 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_32 | 1939 | 26.32 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_33 | 1978 | 26.55 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_34 | 1976 | 26.15 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_35 | 1987 | 26.39 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_36 | 1996 | 26.58 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_37 | 2021 | 22.49 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_38 | 1941 | 24.53 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_39 | 1931 | 24.67 | 1 | 1 | 1 | 1 | 1 |
| FlowCam_40 | 1932 | 29.32 | 1 | 1 | 1 | 1 | 1 |

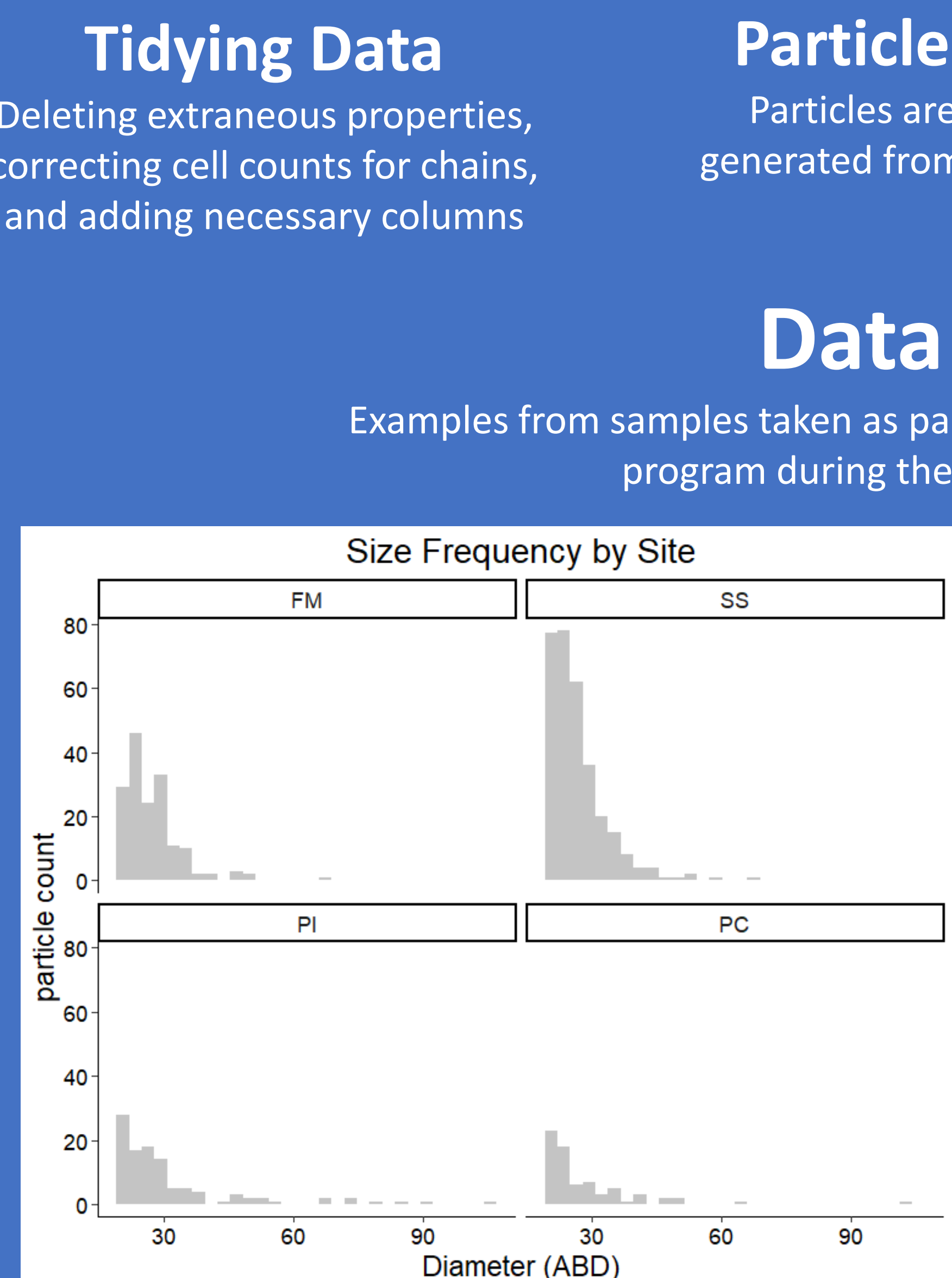


Fig. 2: Histograms illustrating the size spectrum of plankton cells captured at each SWMP site in Diameter (ABD). Due to hardware limitations only cells 20µm-100µm were counted

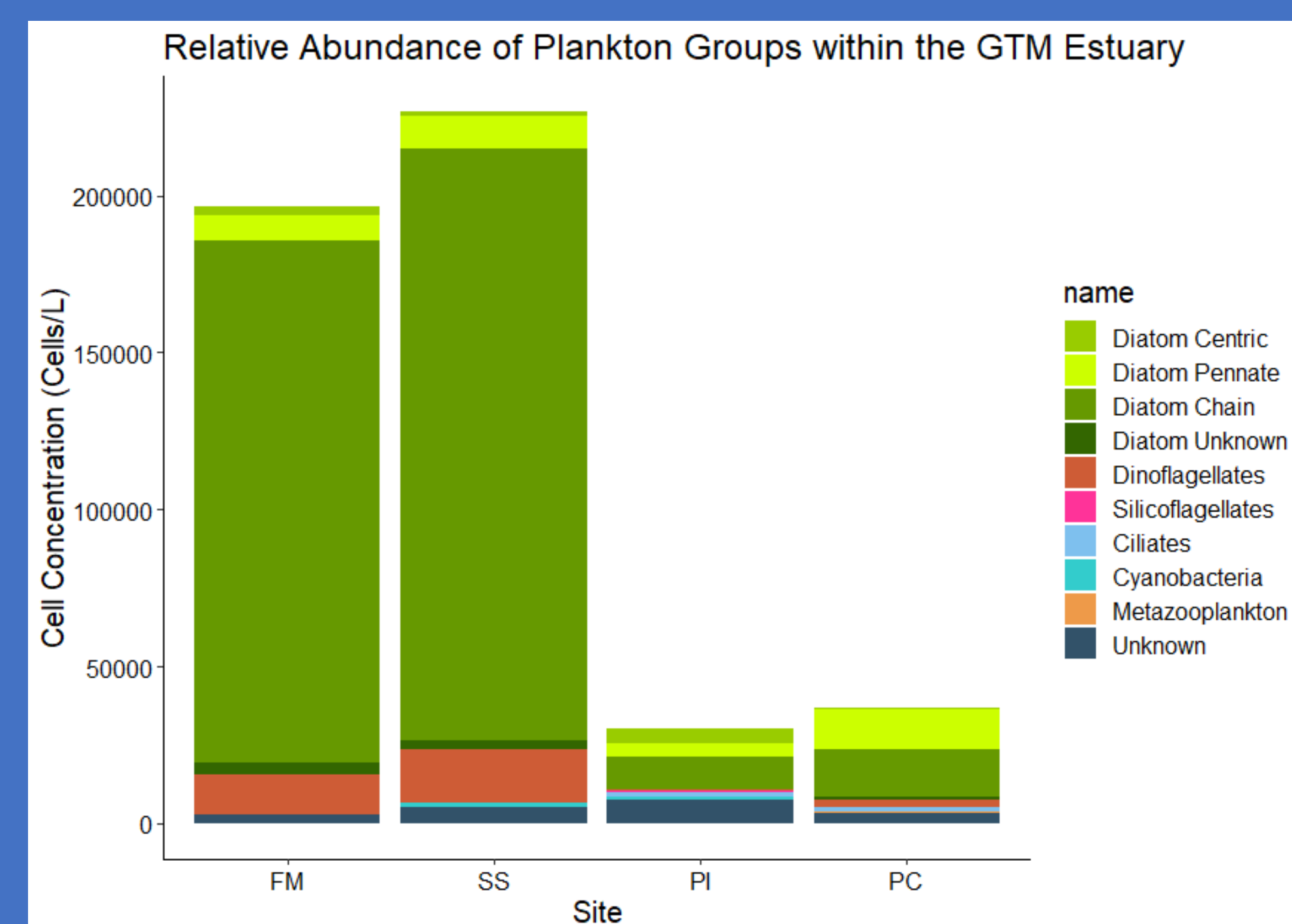


Fig. 3: Relative abundances (cells/L) were calculated at each site for a set of plankton functional types (PFTs). This set of PFTs are groups that exhibit distinct morphological and physiological traits.

A Work in Progress

- Drafting a formal standard operating procedure detailing maintenance and workflows for the FlowCam
- Creating an efficient data pipeline and management plan
- Improving the effectiveness of library generated filters and classification templates
- Exploring the collaborative and educational potential of the FlowCam with volunteers and visiting investigators

Monitoring Goals

- Generate cell counts and size frequencies to characterize plankton community composition within the GTM Estuary
- Develop plankton community characteristics as indicators of changes in water quality and food web dynamics
- Collect actionable and real-time data on the presence of HAB species within the GTM Estuary
- Provide a valuable resource for stakeholders, visiting investigators, and the community

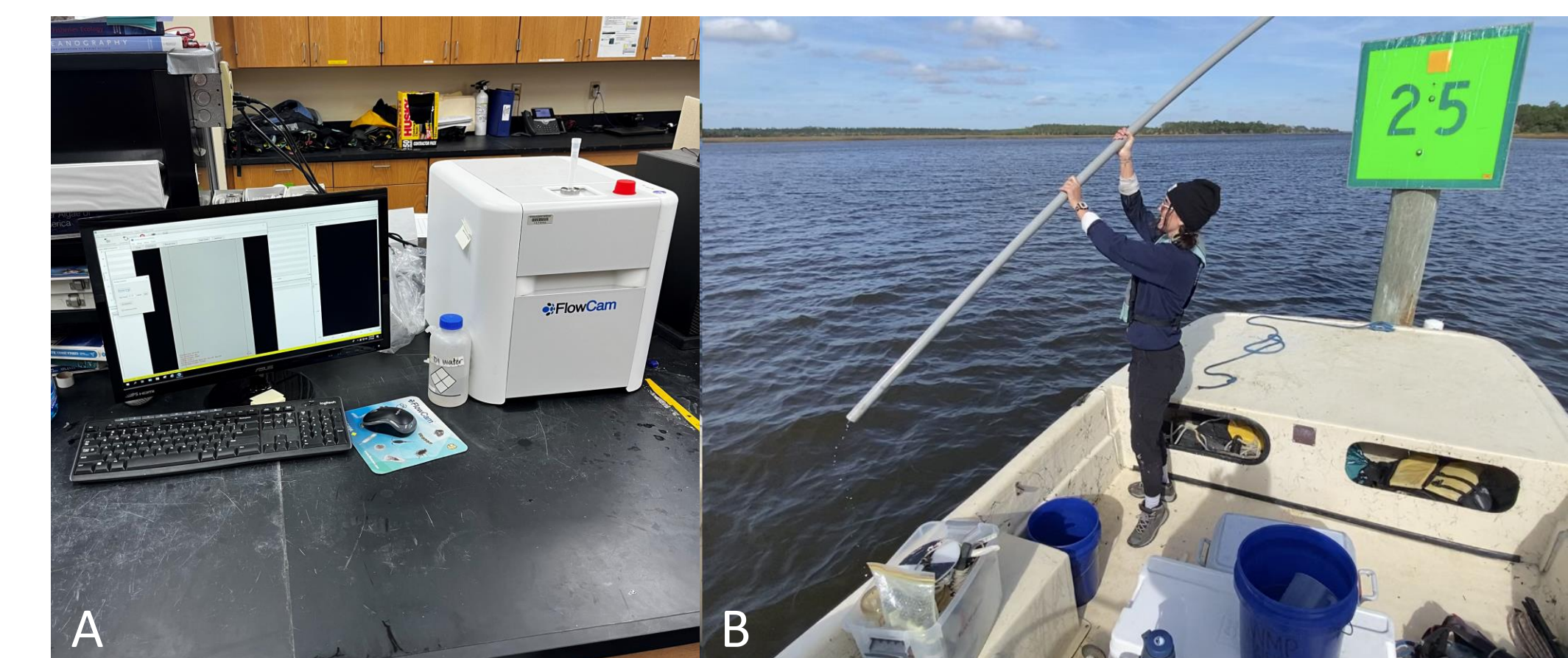


Fig. 4: (A) The GTMNERR's FlowCam 8100 is a stand-alone unit that can easily fit on a desktop or workbench. (B) Technician Megan Howkins collecting water samples during a water quality nutrient sampling run (credit: Silas Tanner)

Significance

- Drastically reduced turnaround times for quantitative plankton data
- Synthesis of plankton and water quality data to identify factors driving shifts in community composition and ecosystem function
- Ability to contribute to global plankton diversity projects with images from the GTM
- FlowCam can be used by visiting investigators to help answer a wide variety of questions.

References

- Naselli-Flores, L., & Padisák, J. (2023). Ecosystem services provided by marine and freshwater phytoplankton. *Hydrobiologia*, 850(12), 2691–2706. <https://doi.org/10.1007/s10750-022-04795-y>
- Chandel, P., Mahajan, D., Thakur, K., Kumar, S., Brar, B., Sharma, D., & Sharma, A. K. (2023). A review on plankton as a bioindicator: A promising tool for monitoring water quality. *World Water Policy*, 1–20. <https://doi.org/10.1002/wwp2.12137>
- Yokogawa Fluid Imaging Technologies Inc. Scarborough, ME. The Ultimate Guide to Flow Imaging Microscopy for Aquatic Life Sciences. <https://info.fluidimaging.com/the-ultimate-guide-to-flow-imaging-microscopy-for-aquatic-life-sciences>. Accessed 2024.