



Environmental (e)DNA-Based Wildlife Monitoring & Human Effluent Quantification Water, Earth & Air







RATORY The Sea Turtle Hospital ENCE at WHITNEY LABORATORY

Duffy Lab, University of Florida







Talk Outline

- Briefly introduce eDNA
- Sea turtle eDNA
- Biodiversity-wide eDNA
- Human eDNA
- Summary

Environmental DNA (eDNA) – wildlife forensics



Organisms continually shed DNA into the environment (skin, hair, bodily fluids etc.)

> Farrell, Whitmore & Duffy. Environmental DNA – how a tool used to detect endangered wildlife ended up helping fight the COVID-19 pandemic. The Conversation, 2021. https://theconversation.com/environmental-dna-how-a-tool-used-to-detect-endangered-wildlife-ended-up-helping-fight-the-covid-19-pandemic-158286

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Organisms continually shed DNA into the environment (skin, hair, bodily fluids etc.)

eDNA – a validated tool. Now being rolled out by federal & state environmental and health management agencies





Our eDNA work:

- Specific wildlife sp.
- Pathogens
- Pan-biodiversity
- Humans



https://www.usatoday.com/story/news/2022/08/26/staggering-dna-advances-could-improve-response-climate-change/10000538002/ https://www.nationalgeographic.com/animals/article/environmental-dna-sheds-light-on-animal-lives-edna https://theconversation.com/environmental-dna-how-a-tool-used-to-detect-endangered-wildlife-ended-up-helping-fight-the-covid-19-pandemic-158286

ANIMALS

Hidden DNA is revealing secrets of animals' lives

New studies of genetic material, or eDNA, shed by plants and animals are providing astonishing biological insights.

NATIONAL

GEOGRAPHIC



from fake payday loans

JSA TODA



Hiring

defies danger signs Job market is solid even as econor

continues to wobble

MAR-A-LAGO SEAL Special maste to review reco











Science's new weapon:



eDNA – coming of age



- Consistently outperforming conventional monitoring approaches for:
 - Native species detection
 - Invasive species monitoring
 - Pathogen monitoring
 - Biodiversity assessments
- Large federally backed initiatives for the adoption of eDNA by managers & conservationists
 - Including eDNA-specific funding to USGS & other agencies in the Bi-partisan Infrastructure Bill (2022)
 - Autonomous samplers & remote sensing
- Health assessments and other metrics still require more traditional survey approaches

Targeted versus untargeted (shotgun) eDNA

- Targeted (qPCR, ddPCR or metabarcoding sequencing)
 - Cost-effective
 - Limited information, more upfront effort required (assay/metabarcode design etc.)
- Shotgun (sequencing)
 - More costly, but vastly more information recovered per sample
 - Simultaneous detection and population genetics/viral variant analysis
 - Can study pan-biodiversity in single sample flora, fauna, microbes & pathogens (e.g. algal blooms, waste/pollution–associated microbes, charismatic megafauna and prey species)
- Combining Targeted & Shotgun can be highly complementary
 - Targeted for large sample numbers & shotgun for deep info. on most crucial samples







eDNA & the GTM NERR

- GTM or GTM-adjacent sites usually make up the majority of our initial test locations
 - Waterways
 - Beaches
 - Air

i.e. in and around the UF Whitney Lab





https://gtmnerr.org/about/













Sand eDNA - Not possible without support of nesting beach patrol volunteers!!

Detection and population genomics of sea turtle species via noninvasive environmental DNA analysis of nesting beach sand tracks and oceanic water

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Beach permit holders & volunteers

ECOLOGY RESOURCES

> Published by Wite



https://onlinelibrary.wiley.com/doi/10.1111/1755-0998.13617

Sea turtle eDNA detectable in nesting beach sand tracks

C. caretta eDNA in wild sand samples collected over 24hrs after single maternal false crawls 0.5 0.4 Concentration (pg/µl 1E-2) 0 0 0.3 œe 0 0 0 0 0 80 0.2 0 0 8 P 0 8 <u></u> **\$**\$6 0.1 do. 8 <u>_____</u> 0.0 0 N S 2 Field negative contro ⁻alse Crawl Cloaca False Nest Surface False Crawl Cloaca False Crawl Cloaca Crawl Flipper **Crawl Flipper** False Crawl Flipper False Nest Deep 1 ⁻alse Nest Surface ⁼alse Nest Surface False Nest Deep False Crawl 1 A False Crawl 1 C False Crawl 1 False False

Turtle eDNA present for at least 24hr after crawl occurred!

Some of our earliest trials were on GTM NERR beaches Scott & Cat Eastman & team

Samples now range from FL, GA, TX, PR, SM

Sand eDNA

• Can we detect eDNA left by a single hatchling (lightly) crawling over sand on its way to the ocean?



Sea turtle eDNA detectable even from a single hatchling crawl



eDNA not just for quantification!

Sea turtle <u>sand eDNA</u> can non-invasively determine geographic origin

Phylogenetic analysis of *C. mydas* and *C. caretta* mitochondrial genomes from sea turtle sand samples and NCBI deposited sequences (whole genome shotgun sequencing)



eDNA for simultaneous pathogen detection

Sand eDNA-based detection of wild tumor-associated pathogen ChHV5

ChHV5 viral genome aligning reads from non-targeted shotgun sequencing of sand eDNA samples (whole genome shotgun sequencing)





ChHV5 viral variant analysis from turtle sand

ChHV5 variant calling from sand eDNA of patient "Archie" (Illumina shotgun sequencing)





of sea turtle genomics quantifi (Submitted for review) Detection, (eDNA) analysis of nesting beach (Submitted for et al. (Z ਰ Ħ Whitmore

0.0050

Can identify viral variants from sand alone! Viral variants: think Covid-19 Alpha, Omicron & Delta

eDNA Metagenomics possible from shotgun sequencing eDNA (pending)



All the major and many of the minor living branches of life are shown on this diagram, but only a few of those that have gone extinct are shown. Example: Dinosaurs - extinct

Pan-species Environmental Metagenomics (NGS-based)

http://evogeneao.s3.amazonaws.com/assets/tree_of_life/tree-of-life_2000.jpg

eDNA – shotgun sequencing interrogated for specific species reptile, mammal, fish & invertebrate



Mark McCauley

Where is eDNA going next?

- Wildlife population genetics from water?
 - Heterogenous mix of species per sample
- Not just what species are present, but where do they come from?

Wildlife population genetics from sea water? (500ml) Human eDNA as proof-of-principle





Also juvenile sea turtle habitat from where we have detected sea turtle eDNA

Human haplogrouping & haplotyping



eDNA can also reveal disease-associated mutations / any alleles of interest in the population



Prominent human cancer-associated genes with or adjacent to deletions detected by gnomAD from Moultrie Creek B water eDNA sample (Oxford Nanopore shotgun long read sequencing)

Same <u>water eDNA</u> Nanopore sample used for haplotyping



Same shotgun eDNA also recovers microbial reads & all other species

Metagenomics taxonomy (human & microbe) high human water (no enrichment Oxford Nanopore shotgun long read sequencing, Moultrie Creek B)



Human eDNA research arose from identifying human eDNA as bycatch in our wildlife studies

Is human eDNA collection possible? Yes

Is human eDNA useful?



Human-specific eDNA assays for rapid and cost-effective wastewater release/septic tank leeching monitoring

- Pollution of aquifers & waterways
- A problem internationally, including NERRs



https://www.surfrider.org/coastalblog/entry/how-do-septic-systemspollute-coastal-watersheds

Effluent Quantification Presumptive high and low human eDNA sites, FL samples

Todd Osborne



https://wusfnews.wusf.usf.edu/environment/2022-09-04/efforts-restore-nature-best-pollution-filters



Human eDNA from intentional water sampling, Florida, human species-specific qPCR assays ZNF285



Human-specific eDNA assays



Human eDNA from intentional water sampling, Florida, human species-specific qPCR assays

Human-specific eDNA assays

Irish river course sampling

• Avoca River, Co. Wicklow





Human eDNA from intentional water sampling, Avoca River, Ireland, <u>LILRB2</u> species-specific (human) qPCR assay



eDNA-based Human Effluent Quantification GTM NERR & Irish validation

- Human-DNA-specific wastewater, septic tank leeching, human fertilizer run-off monitoring
- Those responsible can't claim the source to be wildlife or animal agriculture (as is often the case currently)
- Rapid and cost effective (approx. \$20 per sample), could be coupled with remote monitoring
- Absolute Quantification
 - Can be compared across global sites and large time courses
 - qPCR-based, cost-effective

Human eDNA

- Not just quantification
- Whole genome recovery possible. Utility for:
 - Medical (population-level disease susceptibility)
 - Recover of ancient/contemporary remains & fallen heroes
 - Informing future directions in wildlife/biodiversity eDNA
 - Ethical implications

Human environmental DNA: inadvertent human genomic bycatch and intentional capture raises novel beneficial applications and ethical concerns.

Short title: Promises and dilemmas of human eDNA

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eDNA Summary

• What species are present?



- Where do the present individuals come from? (population genetics of fauna)
- Conservation-relevant genetic loci study (adaptability, disease susceptibility etc.)
- Pathogens (human, livestock and wildlife) & invasive species
- Air eDNA: we <u>have</u> applied to human, sea turtle & pathogens, <u>now</u> working on pan-biodiversity air assessments
- Federal & regulatory support for eDNA to be widely utilized by management agencies



Take-home message eDNA:

- Has HUGE POTENTIAL
- Will be a ROUTINE CONSERVATION & RESILIENCE MANAGEMENT TOOL
- A lot of information in 10g of sand, 500ml water or thin air!











Human eDNA:

My UF lab

start up

Sea Turtle Funding

















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Questions?

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https://www.bbc.com/sport/av/63125522 https://news.ufl.edu/2022/05/sea-turtle-edna/ www.facebook.com/SeaTurtleHospitalatWhitneyLab ww.whitney.ufl.edu/people/current-research-faculty/david-duffy-phd/

Population genetics from water, sand & air? Human eDNA as proof of principle



Voluntary participants – approved by UF ethics board

Environmental DNA (eDNA) – useful for whole sea turtle lifecycle



Human footprint sand Vs 'No human' restricted island sand eDNA







Kurt Foote, Andrew Rich and the NPS staff of the Fort Matanzas National Monument





Supplemental Figure 5





Irish river, town samples

• Avoca River, Co. Wicklow



Mountain tributary

Entering town



Mid-town



Harbour



Beach/mouth

Species-specific qPCR/ddPCR eDNA assays can benefit from reference genomes

- Species—specific assays for (mitochondrial-based):
 - Green (Cm) published (Yetsko et al. 2020, Harper et al. 2020, Yetsko et al. 2021, Farrell et al. 2022)
 - Loggerhead (Cc) published (Farrell et al. 2022)
 - Leatherback (Dc) MS in preparation (Farrell et al.)
 - Kemps (Lk) Active development (Komoroske group)
- Probably wait/need until full references available. Mito. sequences not great for assay design for these species
 - Hawksbill (Ei) need full reference?
 - Olive (Lo) need full reference?
 - Flatback (Nd) need full reference?



Human eDNA from intentional water sampling, Avoca River, Ireland, ZNF285 species-specific (human) qPCR assay



Human eDNA from intentional water sampling, Avoca River, Ireland, <u>LILRB2</u> species-specific (human) qPCR assay



Shotgun sequencing (Illumina) of water and sand eDNA



Problems eDNA can overcome

- Sea turtles are difficult to detect and survey in the wild
 - eDNA no need for visual confirmation or capture of turtles
 - More cost-effective, so can survey more areas
- Population genetic data for conservation and research requires invasive blood/tissue sampling, and egg sacrifice is used
 - eDNA means no disturbance of individuals, especially females during nesting
 - Samples can be taken from sand hours after the nesting event
 - So don't need to see nesting, can get samples the next morning



Sea turtle eDNA detectable in nesting beach sand tracks



Sea turtle population genetics from sea water? Human eDNA as proof of principle



Native American





